

FGF and ERK signaling coordinately regulate mineralization-related genes and play essential roles in osteocyte differentiation

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Abstract To examine the roles of FGF and ERK MAPK signaling in osteocyte differentiation and function, we performed microarray analyses using the osteocyte cell line MLO-Y4. This experiment identified a number of mineralization-related genes that were regulated by FGF2 in an ERK MAPK-dependent manner. Real-time PCR analysis indicated that FGF2 upregulates *Ank*, *Enpp1*, *Mgp*, *Slc20a1*, and *Dmp1* in MLO-Y4 cells. Consistent with this observation, the selective FGF receptor inhibitor PD173074 decreased *Ank*, *Enpp1*, *Slc20a1*, and *Dmp1* mRNA expression in mouse calvaria in organ culture. Since *Dmp1* plays a central role in osteocyte differentiation and mineral homeostasis, we further analyzed FGF regulation of *Dmp1*. Similar to FGF2, FGF23 upregulated *Dmp1* expression in MLO-Y4 cells in the presence of

Klotho. Furthermore, increased extracellular phosphate levels partially inhibited FGF2-induced upregulation of *Dmp1* mRNA expression, suggesting a coordinated regulation of *Dmp1* expression by FGF signaling and extracellular phosphate. In MLO-Y4 osteocytes and in MC3T3E1 and primary calvaria osteoblasts, U0126 strongly inhibited both basal expression of *Dmp1* mRNA and FGF2-induced upregulation. Consistent with the in vitro observations, real-time PCR and immunohistochemical analysis showed a strong decrease in *Dmp1* expression in the skeletal elements of *ERK1*^{-/-}; *ERK2*^{flx/flx}; *Prx1-Cre* mice. Furthermore, scanning electron microscopic analysis revealed that no osteocytes with characteristic dendritic processes develop in the limbs of *ERK1*^{-/-}; *ERK2*^{flx/flx}; *Prx1-Cre* mice. Collectively, our observations indicate that FGF signaling coordinately regulates mineralization-related genes in the osteoblast lineage and that ERK signaling is essential for *Dmp1* expression and osteocyte differentiation.

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Introduction

Osteoblasts arise from undifferentiated mesenchymal cells and undergo sequential steps of differentiation. Osteoblasts lay down osteoid matrix, coordinate its mineralization, and subsequently become embedded in bone matrix as osteocytes. Numerous studies have indicated that the differentiation and function of osteoblasts are under the control of various systemic hormones and local cytokines and growth factors. Some of the important regulators of osteoblast differentiation are fibroblast growth factors (FGFs). The

inactivation of *Fgf2* in mice causes decreased bone mass and decreased bone formation [1]. Furthermore, the genetic inactivation of *Fgfr2* causes reduced osteoblast proliferation and increased osteopenia, whereas *Fgfr1* has been implicated in stage-dependent regulation of osteoblast proliferation and differentiation [2, 3]. In humans, activating mutations in FGFR2 cause syndromic craniosynostosis, such as Apert and Crouzon syndromes [4]. These observations indicate that FGF signaling plays an essential role in the control of osteoblast differentiation and bone formation.

FGF signaling is mediated by various intracellular signaling pathways including protein kinase C, phospholipase C- γ , and the extracellular signal-regulated kinase (ERK) mitogen-activated protein kinase (MAPK) pathway [5]. Studies from our laboratory as well as other investigators have shown that the ERK MAPK pathway plays an essential role in mediating FGF signaling in skeletal cells [6, 7]. Our studies using genetically engineered mouse models have indicated that ERK1 and ERK2 play an essential role in osteoblast and chondrocyte differentiation [8]. The genetic inactivation of *ERK1* and *ERK2* in undifferentiated mesenchymal cells leads to severe impairment of osteoblast differentiation and bone formation. In addition, a number of missense-activating mutations in HRAS, KRAS, BRAF, MEK1, and MEK2 have been identified in Costello, Noonan, LEOPARD, and cardio-facio-cutaneous syndromes, highlighting the importance of the ERK MAPK pathway in human skeletal development [9, 10].

While numerous *in vivo* and *in vitro* studies have examined the mechanisms of osteoblast differentiation, relatively little is known about the transition from osteoblasts to osteocytes. Osteocytes are non-proliferative, terminally differentiated cells in the osteoblast lineage, and the osteocyte is the most abundant cell type in bone. Recent studies have indicated that osteocytes function as a mechanosensor in the bone, coordinate bone homeostasis, and secrete dentin matrix protein 1 (Dmp1) and FGF23 to regulate phosphate homeostasis [11–14]. Dmp1 is a non-collagenous protein expressed in the osteoblast lineage, which is most abundantly expressed in osteocytes. Dmp1 has been shown to play a critical role in osteocyte differentiation [11, 15]. *Dmp1*-null mice show defective osteoblast-to-osteocyte maturation and defects in mineralization. Furthermore, DMP1 mutations in humans cause autosomal recessive hypophosphatemic rickets (ARHR) and osteomalacia. Interestingly, both *Dmp1*-null mice and patients with DMP1 mutations show increased FGF23 expression in osteocytes and elevated systemic FGF23 levels, indicating that DMP1 acts upstream of FGF23 and regulates phosphate metabolism.

To investigate the role of FGF and ERK MAPK signaling in the late stages of osteoblast differentiation, we

examined the effects of FGF2 and a MEK inhibitor, U0126, in MLO-Y4 osteocytes, an immortalized cell line that maintains the characteristics of osteocytes [16]. We show here that FGF2 coordinately regulates a number of mineralization-related genes in an ERK MAPK-dependent manner in MLO-Y4 cells. Our *in vivo* and *in vitro* observations indicate that Dmp1 expression in osteoblasts and osteocytes requires FGF and ERK signaling. Furthermore, scanning electron microscopic (SEM) analysis shows that no osteocytes with characteristic dendritic processes develop in the absence of ERK1 and ERK2. These observations indicate that FGF and ERK signaling plays an essential role in osteocyte differentiation and mineral homeostasis.

Materials and methods

Cell and calvaria organ culture

MLO-Y4 cells were cultured in α -MEM containing 2.5% FCS and 2.5% calf serum. MC3T3-E1 cells were cultured in α -MEM containing 10% FCS. Primary calvaria osteoblasts were isolated from neonatal mouse calvaria after sequential digestion with 0.1% collagenase and 0.2% dispase as described previously [17]. The cells were grown in α -MEM, 10% FCS, and the medium was further supplemented with 5 mM β -glycerophosphate and 100 μ g/ml ascorbic acid upon reaching confluence. For embryonic calvaria organ culture, calvariae were isolated from wild-type embryos at 18.5 days post-gestation. Each calvaria was individually placed in a well of a 48-well plate containing 200 μ l α -MEM supplemented with 10% FCS, 50 μ g/ml ascorbic acid, 1 mM β -glycerophosphate, and penicillin/streptomycin. Cultures were maintained at 37°C in a humidified incubator with 5% CO₂ in air. Recombinant human FGF2, mouse FGF23, and mouse Klotho were purchased from R&D. MEK1/MEK2 inhibitor U0126 and FGF receptor inhibitor PD173074 were purchased from Promega and Tocris Bioscience, respectively. Cycloheximide and heparin were purchased from Sigma.

To determine the effect of extracellular inorganic phosphate concentration on MLO-Y4 cells, cells were incubated in α -MEM containing 0.25% FCS and 0.25% calf serum for 16 h 40 min and treated for 8 h with FGF2 and inorganic phosphate, which was added as a mixture of NaH₂PO₄ and Na₂HPO₄ (pH 7.2).

To examine the effects of FGF2 on mineralization, MLO-Y4 cells and primary calvaria osteoblasts were plated in 24-well plates, and the medium was supplemented with 5 mM of β -glycerophosphate and 100 μ g/ml of ascorbic acid with or without 10 ng/ml FGF2 upon reaching confluence. The medium was changed every

3 days. Mineralization was evaluated by alizarin red staining at 10 days after the induction of mineralization. Cells were fixed in 70% ethanol for 15 min and stained with 1% alizarin red solution.

Mutant animals

The institutional animal care and use committee of Case Western Reserve University approved all animal procedures. All animal care and use were performed in accordance with the institutional animal use methods and policies. *ERK1*-null mice and mice with the floxed allele of *ERK2* were described previously [18, 19]. *Prx1-Cre* transgenic mice were obtained from James Martin [20]. *ERK1*^{-/-}; *ERK2*^{lox/lox}; *Prx1-Cre* embryos were generated as described [8]. Mice with the floxed allele of *Fgfr2* were purchased from the Jackson Laboratory [2].

RNA extraction and semi-quantitative and real-time PCR

Total RNA was extracted from cells and mouse skeletal elements using the RNeasy kit (Qiagen). RNA was reverse-transcribed to cDNA with the High Capacity cDNA Archive Kit (Applied Biosystems). Real-time PCR was performed on the Applied Biosystems 7500 Real-time PCR detection system. All the real-time PCR analyses were done in triplicate or quadruplicate. TaqMan probe sets were designed and synthesized by Applied Biosystems [*Dmp1* Mm00803833_g1, *Erk2* Mm00442479_m1, *Enpp1* Mm00501097_m1, *Mgp* Mm00485009_m1, *Ank* Mm00445040_m1, *Slc20a1* Mm00489378_m1, *Fgf23* Mm00445621_m1, *Gapdh* 4352932E, *sclerostin (Sost)* Mm00470479_m1, and *klotho* Mm00502002_m1]. To compare gene expression levels, the comparative cycle threshold (Ct) method was used. *Gapdh* was used as an endogenous control to correct for potential variation in RNA loading or in efficiency of amplification. Semi-quantitative PCR was performed on the Applied Biosystems GeneAmp PCR system 9700 using the following primer sets: *Dmp1* (forward) 5'-AGATCCCTCTTCGAGAAGTTCGCT-3' and (reverse) 5'-TTCTGATGACTCACTGTTTCGTGGGTG-3', PCR product: 242 bp; *Fgfr1*, (forward) 5'-TGGAGTTCA TGTGTAAGGTG-3' and (reverse) 5'-ATAAAGAGG ACCATCCTGTG-3', PCR product: 856 bp; *Fgfr2*, (forward) 5'-AAATACCAAATCTCCCAACC-3' and (reverse) 5'-GCCGCTTCTCCATCTTCT-3', PCR product: 373 bp; *Fgfr3*, (forward) 5'-ACTGTAAGTCAAGACTGCAGG-3' and (reverse) 5'-GTCCTTGTCAGTCGCATCAT-3', PCR product: 635 bp; *Fgfr4*, (forward) 5'-TACAGTGGCTG AACACGTCGTC-3' and (reverse) 5'-ACAAGC

AGAACCAGTGAGCCTGAT-3', PCR product: 304 bp; *Gapdh* (forward) 5'-ACCACAGTCCATGCCATCAC-3' and (reverse) 5'-TCCACCCTGTTGCTGTA-3', PCR product: 452 bp. cDNA was amplified 28–35 cycles, and the band intensity was compared while the band intensity and cycle numbers were linear.

Microarray

Microarray analysis using Affymetrix Mouse Genome 430 2.0 Array was performed at the Gene Expression and Genotyping Core Facility of the Case Comprehensive Cancer Center at Case Western Reserve University. The differentially expressed genes were identified using the Affymetrix GCOS software and the Ariadne Pathway Studio software.

Western blot analysis

Phosphorylation of ERK1 and ERK2 was determined by Western blot analysis using anti-ERK1 antibody (Santa Cruz Biotechnology, sc-94) and anti-phospho-p44/42 MAP kinase (Thr202/Tyr204) antibody (Cell Signaling Technology, #9101S). For Western blot detection of Dmp1, MLO-Y4 cells in 100-mm dishes were serum starved and treated with 10 ng/ml FGF2 for 21 h. Culture supernatant was concentrated from 6 ml to 135 µl using centrifugal filter units (Millipore; Amicon Ultra-15) and supplemented with proteinase inhibitor cocktail (Thermo Scientific). Total cell lysates were prepared by lysing the cells in 20 mM Tris-HCl (pH 8.0), 150 mM NaCl, 1% Triton X-100, 1 mM sodium orthovanadate, 1 mM sodium fluoride, and 1 mM β-glycerophosphate, supplemented with proteinase inhibitor cocktail. Twenty microliters of concentrated medium or 40 µg of total cellular protein was separated by 10% or 4–15% sodium dodecyl sulfate-polyacrylamide gel electrophoresis and electrophoretically transferred to polyvinylidene difluoride filters (Millipore). The filters were blocked in StartingBlock T20 Blocking Buffers (Thermo Scientific) containing 0.05% Tween-20 and then incubated with anti-mouse Dmp-1 antibody (R&D, AF4386) at 0.4 µg/ml. Filters were then incubated with the horseradish peroxidase-conjugated donkey anti-sheep IgG antibody (SantaCruz), and the signal was detected by enhanced chemiluminescence (Thermo Scientific). The signal was quantitated using ImageJ software (NIH, Bethesda, MD). Dmp1 protein levels in the supernatant were normalized by the total cellular protein as determined by bicinchoninic acid protein assay kit (Thermo Scientific). As an internal control, the expression of β-actin protein was evaluated using the anti-β-actin antibody (Cell Signaling Technology, #4967).

Histology

For von Kossa staining, bone samples were fixed in 10% formalin and embedded in paraffin. Seven-micron sections were stained with 5% silver nitrate solution for 5 min, fixed with 5% sodium thiosulfate for 3 min, and counterstained with nuclear fast red (KPL, Contrast RED). For Goldner's trichrome staining, bone tissues were embedded in methyl methacrylate, and sections were stained using a standard protocol. For immunohistochemical detection of Dmp1, bone samples were fixed in 10% formalin, demineralized in 0.5 M EDTA, and embedded in paraffin. Seven-micron sections were deparaffinized, and endogenous peroxidase activity was blocked with 3% H₂O₂ in methanol for 10 min. Then the sections were blocked with 10% normal donkey serum in phosphate buffer saline for 30 min. Sections were reacted with anti-mouse Dmp-1 antibody (R&D) for 2 h at room temperature. For control, anti-mouse Dmp-1 antibody was either omitted or replaced by normal sheep IgG at the same concentration. The sections were further incubated with the secondary antibody (donkey anti-sheep IgG conjugated to biotin) for 30 min, and reacted with streptavidin horseradish-peroxidase (Biocare) for 15 min, followed by incubation with diaminobenzidine (Vector Laboratories: ImmPACT DAB). Sections were counterstained with hematoxylin. Samples were photographed using a Leica DM 6000B microscope, Leica DC500 digital camera, and Leica Application Suite 1.3 software.

Scanning electron microscopy

To image the osteocyte lacunar-canalicular system, acid-etched, resin-casted samples were analyzed by SEM [21]. Bone samples were first fixed in 70% ethanol, embedded in methyl methacrylate, and sliced with a tungsten carbide knife. The surface of the specimens was polished sequentially using a wet 1,200 grit Carbimet 2 Abrasive Disc (P2500, Buehler) and Microcloth (Buehler) with 1.0, 0.25, and 0.05 μ m Metadi Supreme Diamond suspension (Buehler). The polished surface was acid-etched in 9% phosphoric acid for 1 min and immersed in sodium hypochlorite (bleach) for 5 min. Specimens were sputter coated with 3–5 nm of palladium using Denton Desk IV. Scanning electron microscope images were collected using the Helios Nanolab-650 Dual Beam FIB system (FEI) or Nova Nano-lab 200 (FEI) with accelerating voltage of 2–3 kV.

Measurement of serum phosphate concentration

To determine serum phosphate concentration, blood was collected from control and *ERK1*^{-/-}; *ERK2*^{flx/flx}; *Pxx1-Cre* mice at P0. Then serum was separated, and phosphate

concentration was measured using the phosphate assay kit (Stanbio).

Statistical analysis of data

Statistical analysis was performed using Analyse-it (Analyse-it Software). A one-way ANOVA analysis followed by Fisher's least significant difference (LSD) for post-hoc analysis was used to compare the four groups of different treatments. A *t* test was applied to compare the control group and PD173074 treatment group. *p* values <0.05 were considered statistically significant.

Results

FGF receptor expression and FGF2-induced ERK activation in MLO-Y4 cells

To identify FGF receptors that mediate FGF signaling in MLO-Y4 cells and primary calvaria osteoblasts, we first examined the expression of *Fgfr1*, *Fgfr2*, *Fgfr3*, and *Fgfr4* by semi-quantitative RT-PCR. While primary calvaria osteoblasts expressed all of the four FGF receptors, only *Fgfr1* and *Fgfr3* mRNAs were detected in MLO-Y4 cells (Fig. 1a).

We next examined activation of the ERK MAPK pathway by FGF2 and its inhibition by the MEK inhibitor U0126 in MLO-Y4 cells. Western blot analysis using the phospho-specific antibody indicated that FGF2 induces ERK phosphorylation at Thr202 and Tyr204, and that U0126 strongly inhibits FGF2-induced ERK phosphorylation (Fig. 1b).

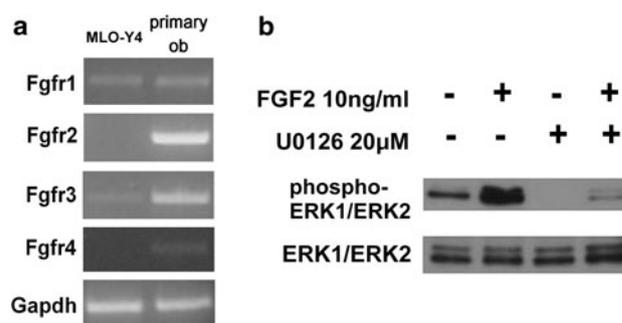


Fig. 1 **a** FGF receptor expression in MLO-Y4 cells and primary calvaria osteoblasts (primary ob). The expression of *Fgfr1*, *Fgfr2*, *Fgfr3* and *Fgfr4* was examined by semi-quantitative RT-PCR. **b** Western blot analysis showing the activation of the ERK MAPK pathway by FGF2 and its inhibition by U0126. MLO-Y4 cells were treated with U0126 or its solvent DMSO for 45 min followed by FGF2 treatment for 120 min. Total cell lysates were subjected to Western blot analysis. The *bottom panel* shows total ERK1/2 expression and *upper panel* shows phosphorylated ERK1/2

FGF2 regulates mineralization-related genes in MLO-Y4 cells through the ERK MAPK pathway

To investigate the roles of the ERK MAPK pathway and FGF signaling in the late stages of osteoblast differentiation, we performed microarray analysis using the osteocyte cell line MLO-Y4. MLO-Y4 cells were treated with 10 ng/ml FGF2 in the presence or absence of 20 μ M U0126. RNA was extracted 3 and 8 h after the treatment and subjected to microarray analysis. We identified 172 genes that were reproducibly increased or decreased at least twofold at 3 h after FGF2 treatment in two independent experiments (Supplementary Table 1). We also identified 434 genes that were reproducibly increased or decreased at least twofold at 8 h after FGF2 treatment. U0126 treatment reduced the number of FGF2-regulated genes to 25 and 85 at 3 and 8 h, respectively. Among genes that were differentially regulated by FGF2, at least four mineralization-related genes, *Ank*, *Dmp1*, *Mgp*, and *Slc20a1*, were upregulated by FGF2 in MLO-Y4 cells (Table 1). We also found that *Enpp1* was downregulated twofold by U0126 at 8 h after treatment in one of the two experiments.

The regulation of the five mineralization-related genes, *Ank*, *Dmp1*, *Mgp*, *Slc20a1*, and *Enpp1*, was further

confirmed by real-time PCR. FGF2 upregulated all of the five mineralization-related genes two- to sixfold after treatment, and this FGF2-induced upregulation was inhibited by U0126 ($p < 0.01$) (Fig. 2). In the presence of U0126, FGF2 did not significantly increase *Ank*, *Dmp1*, *Mgp*, and *Slc20a1* expression, strongly suggesting that the FGF2-induced upregulation was mediated by the ERK MAPK pathway. Furthermore, U0126 strongly inhibited basal expression of *Dmp1* mRNA ($p < 0.05$). *Dmp1* mRNA expression decreased to 16% of control at 8 h after U0126 treatment. These results suggest that *Dmp1* expression depends on ERK MAPK signaling.

PD173074 inhibits mineralization-related genes in mouse calvaria

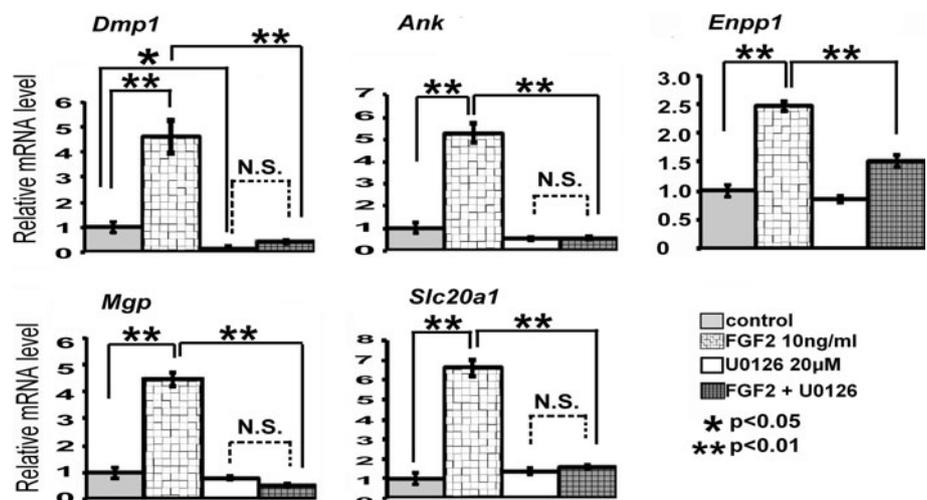
To address the role of FGF signaling in a more physiological context, we treated the whole neonatal calvaria with PD173074 in an organ culture system. PD173074 is a selective FGF receptor inhibitor that has been shown to inhibit all four FGF receptors [22–24]. The neonatal calvaria was treated with 1 μ M of PD173074 for 24 h. The effects of PD173074 on various other kinases have been shown to be minimal at this concentration [23]. PD173074

Table 1 Five mineralization-related genes were identified by microarray analysis of FGF2 and U0126-treated MLO-Y4 cells

Gene		Fold changes		
Symbol	Name	FGF2 vs. control	U0126 vs. control	FGF2 + U0126 vs. control
<i>Dmp1</i>	Dentin matrix protein 1	7.07	−11.1	−1.08
<i>Ank</i>	Progressive ankylosis	3.56	−1.96	−1.12
<i>Mgp</i>	Matrix Gla protein	2.26	−1.38	−1.45
<i>Slc20a1</i>	Solute carrier family 20, member 1	6.0	1.39	2.49
<i>Enpp1</i>	Ectonucleotide pyrophosphatase/phosphodiesterase 1	1.0	−1.71	−1.21

The cells were treated with 10 ng/ml FGF2 in the presence or absence of 20 μ M U0126 for 8 h. The table shows the average fold changes of two independent experiments

Fig. 2 Five mineralization-related genes were upregulated by FGF2 in MLO-Y4 cells, and the FGF2-induced upregulation was inhibited by U0126. The cells were treated with 10 ng/ml FGF2 in the presence or absence of 20 μ M U0126. RNA was extracted at 3 h after FGF2 treatment for *Enpp1* and at 8 h for *Dmp1*, *Ank*, *Mgp*, and *Slc20a1*. mRNA expression levels were examined by real-time PCR in triplicate. Data represent mean \pm standard deviations. The figure presents data from one of two experiments with similar results. *N.S.* not significant



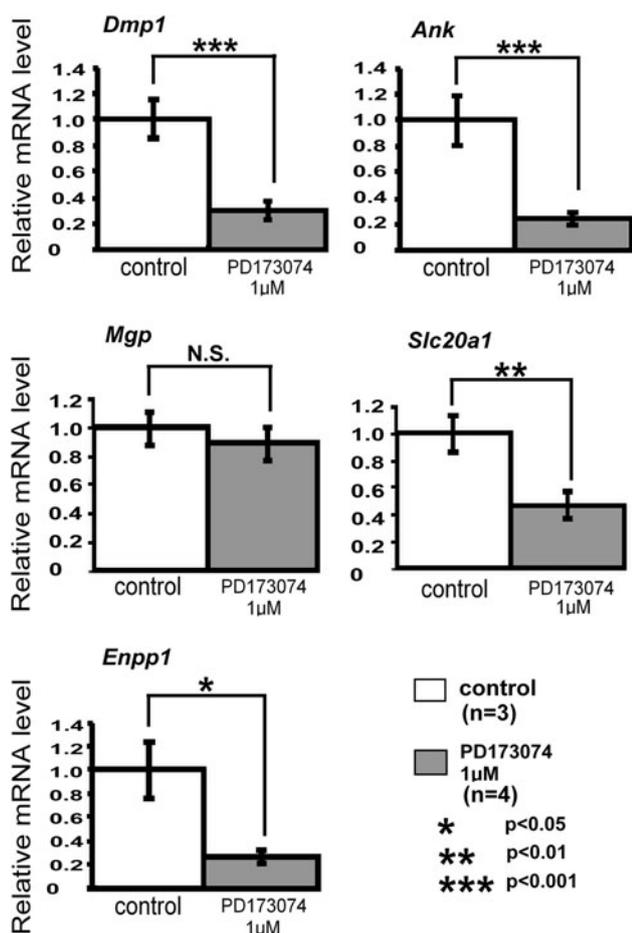


Fig. 3 PD173074 inhibits expression of *Dmp1*, *Ank*, *Slc20a1*, and *Enpp1* in mouse calvaria in organ culture. Calvariae isolated from wild-type embryos at 18.5 days post-gestation were maintained in organ culture and treated with the selective FGF receptor inhibitor PD173074 at 1 µM for 24 h. Expression of mineralization-related genes, *Dmp1*, *Ank*, *Mgp*, *Slc20a1*, and *Enpp1*, was examined by real-time PCR. Real-time PCR was done in triplicate for each calvaria sample. Data represent mean ± standard deviations of 3 control calvariae and 4 calvariae treated with PD173074. The figure presents data from one of two experiments with similar results. **p* < 0.05, ***p* < 0.01, ****p* < 0.001

inhibited *Dmp1*, *Ank*, *Enpp1*, and *Slc20a1* expression by 70, 76, 74, and 53% of the control, respectively, while *Mgp* expression was unaffected (Fig. 3). These findings suggest that *Dmp1*, *Ank*, *Enpp1*, and *Slc20a1* expression depends on FGF signaling.

FGF2 inhibits mineralization of primary calvaria osteoblasts

To examine the overall effects of FGF2 on mineralization, mineralization was induced in MLO-Y4 cells and primary calvaria osteoblasts in the presence or absence of FGF2. FGF2 treatment for 10 days resulted in reduced mineralization of primary calvaria osteoblasts as shown by alizarin

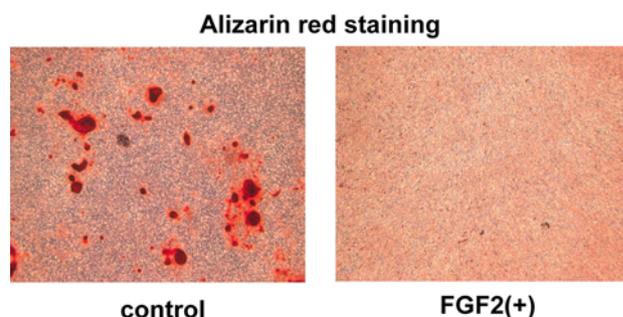


Fig. 4 FGF2 inhibited matrix mineralization of primary calvaria osteoblasts. Mineralization was induced in confluent primary osteoblast culture for 10 days by supplementing the medium with 5 mM β-glycerophosphate and 100 µg/ml ascorbic acid in the presence or absence of 10 ng/ml FGF2. Matrix mineralization was evaluated by alizarin red staining

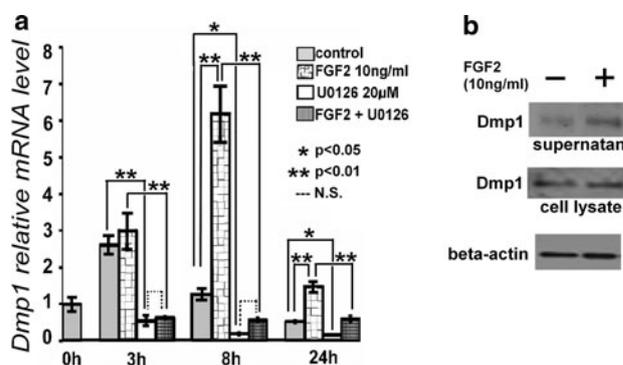


Fig. 5 FGF2 increases *Dmp1* mRNA and protein levels in MLO-Y4 cells. **a** Time-dependent increase of *Dmp1* mRNA after FGF2 treatment. The cells were treated with 10 ng/ml FGF2 in the presence or absence of 20 µM U0126. RNA was isolated at 3, 8 and 24 h after FGF2 treatment. *Dmp1* mRNA levels were measured by real-time PCR in triplicate. Data represent mean ± standard deviations. **b** FGF2 increases *Dmp1* protein levels in the culture supernatant of MLO-Y4 cells, while *Dmp1* protein levels in total cell lysates remain unchanged. MLO-Y4 cells were treated with 10 ng/ml FGF2 in the absence of serum for 21 h. Culture supernatant was concentrated using centrifugal filter units and subjected to Western blot analysis. The bottom panel shows β-actin protein levels in total cell lysates as a loading control. The figure presents data from one of four experiments with similar results. N.S. not significant

red staining (Fig. 4). In contrast, MLO-Y4 cells did not mineralize their surrounding matrices as described previously [25].

FGF2 increases *Dmp1* mRNA and protein levels in MLO-Y4 cells

Since *Dmp1* has been implicated in osteocyte differentiation, we focused our study on the regulation of *Dmp1*. We examined the time course of *Dmp1* mRNA upregulation after FGF2 treatment. FGF2-induced *Dmp1* expression reached its maximum at 8 h, and *Dmp1* mRNA levels decreased thereafter (Fig. 5a). We further confirmed

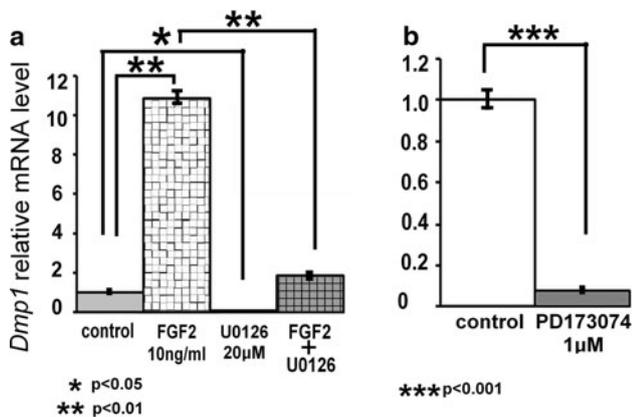


Fig. 6 FGF2 upregulates and PD173074 inhibits *Dmp1* expression in primary calvaria osteoblasts. **a** Primary calvaria osteoblasts were treated with 10 ng/ml FGF2 in the presence or absence of 20 μM U0126. RNA was isolated at 7 h after FGF2 treatment. **b** Primary calvaria osteoblasts were treated with the selective FGF receptor inhibitor PD173074 at 1 μM. RNA was isolated at 31 h after PD173074 treatment. *Dmp1* mRNA levels were examined by real-time PCR in triplicate. Data represent mean ± standard deviations. The figure presents data from one of two experiments with similar results

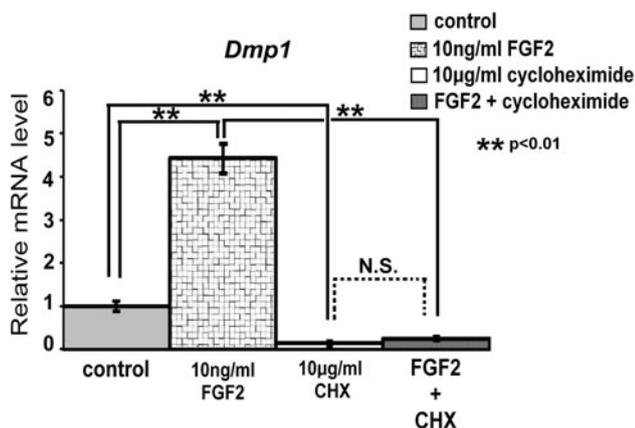


Fig. 7 Cycloheximide inhibits FGF2-induced upregulation of *Dmp1* expression. MLO-Y4 cells were treated with 10 ng/ml FGF2 in the presence or absence of 10 μg/ml cycloheximide (CHX). RNA was extracted at 7 h after FGF2 treatment, and *Dmp1* mRNA expression was examined by real-time PCR in triplicate. Data represent mean ± standard deviations. The figure presents data from one of two experiments with similar results. *N.S.* not significant

FGF2-induced *Dmp1* upregulation at the protein level by Western blot analysis. It has been shown that full-length *Dmp1* is proteolytically cleaved to form two fragments of approximately 37 and 57 kDa [26]. The 57-kDa C-terminal fragment has been shown to regulate osteocyte maturation and mineralization [15]. *Dmp1* protein was detected as a single band at 57 kDa in the culture supernatant of MLO-Y4 cells (Fig. 5b). FGF2 treatment increased *Dmp1* protein levels in the supernatant two- to sixfold, consistent with

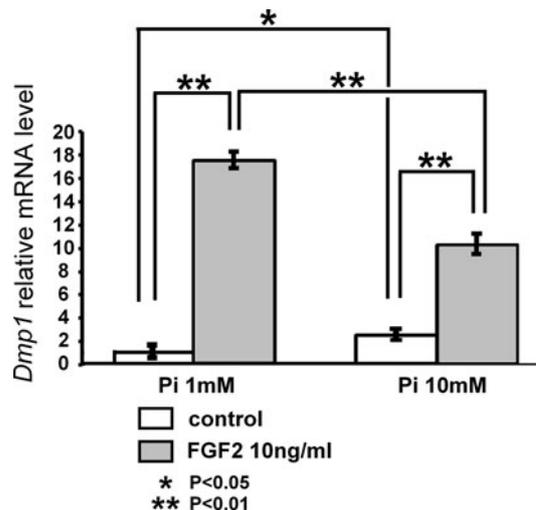


Fig. 8 Increased phosphate concentration diminishes the magnitude of FGF2 effects on *Dmp1* regulation in MLO-Y4 cells. Cells were cultured in α-MEM containing 0.25% FCS and 0.25% calf serum, and treated with 10 ng/ml FGF2 in the presence of 1 or 10 mM phosphate. RNA was extracted at 8 h after FGF2 treatment, and *Dmp1* mRNA expression was examined by real-time PCR in triplicate. Data represent mean ± standard deviations. The figure presents data from one of three experiments with similar results. **p* < 0.05, ***p* < 0.01

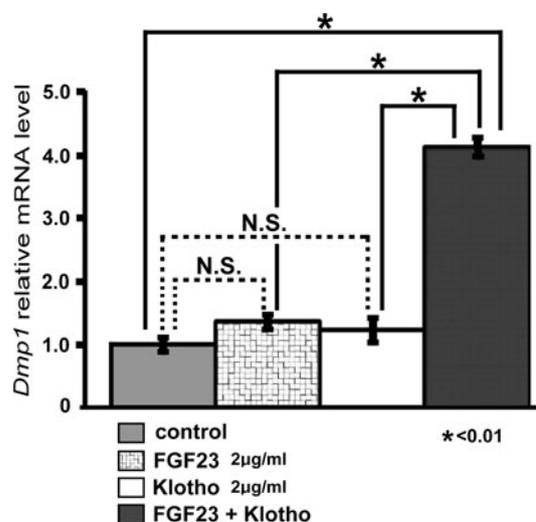


Fig. 9 FGF23 upregulates *Dmp1* expression in MLO-Y4 cells in the presence of Klotho. Cells were incubated in α-MEM containing 0.25% FCS and 0.25% calf serum for 17 h and treated with 2 μg/ml FGF23 and 2 μg/ml Klotho either alone or in combination in the presence of 50 μg/ml heparin. RNA was extracted at 8 h after the treatment, and *Dmp1* mRNA expression was examined by real-time PCR in triplicate. Data represent mean ± standard deviations. The figure presents data from one of three experiments with similar results. **p* < 0.01. *N.S.* not significant

FGF2 upregulation of *Dmp1* mRNA expression. We did not observe an obvious increase in *Dmp1* protein levels in total cell lysates, suggesting that *Dmp1* protein does not accumulate inside the cells (Fig. 5b).

FGF2 regulates *Dmp1* expression in primary calvaria osteoblasts

To examine the generality of FGF2-induced upregulation of *Dmp1* mRNA in the osteoblast lineage, we further examined the effects of FGF2 in primary calvaria osteoblasts and preosteoblastic cell line MC3T3E1. Similar to MLO-Y4 osteocytes, FGF2 increased *Dmp1* mRNA expression in primary calvaria osteoblasts and MC3T3E1 cells, and U0126 strongly inhibited basal *Dmp1* expression and FGF2-induced upregulation (Fig. 6a and data not shown). In addition, the treatment of primary calvaria osteoblasts with selective FGF receptor inhibitor PD173074 strongly inhibited *Dmp1* expression ($p < 0.001$) (Fig. 6b).

FGF2-induced upregulation of *Dmp1* expression requires new protein synthesis

To further examine the mechanism of FGF2-induced upregulation of *Dmp1* expression, we treated MLO-Y4 cells with 10 ng/ml FGF2 in the absence and presence of 10 μ g/ml cycloheximide to block protein synthesis. FGF2-induced *Dmp1* expression was significantly inhibited in the presence of cycloheximide ($p < 0.01$), indicating that FGF2-induced upregulation of *Dmp1* expression requires new protein synthesis (Fig. 7). Interestingly, cycloheximide treatment alone strongly inhibited *Dmp1* expression, suggesting that continued protein synthesis is required for basal *Dmp1* expression.

Inorganic phosphate levels modulate FGF2 regulation of *Dmp1* expression in MLO-Y4 cells

Since *Dmp1* is implicated in phosphate homeostasis, we tested the effects of extracellular phosphate concentration on the regulation of *Dmp1* mRNA expression by FGF2. MLO-Y4 cells were cultured in α -MEM containing 0.25% FCS and 0.25% calf serum and treated with 10 ng/ml FGF2 and sodium phosphate to achieve final phosphate concentrations of 1 and 10 mM. At phosphate concentration of 10 mM, *Dmp1* mRNA levels induced by FGF2 were diminished by 40% compared to the cells cultured at low phosphate concentration of 1 mM (Fig. 8). On the other hand, phosphate treatment alone increased *Dmp1* mRNA expression levels in the absence of FGF2. Overall, the magnitude of FGF2 effects on *Dmp1* regulation is diminished by increased phosphate concentration.

FGF23 increases *Dmp1* expression in MLO-Y4 cells in the presence of Klotho

Since osteocytes produce FGF23, we further examined the effects of FGF23 on *Dmp1* expression. MLO-Y4 cells were

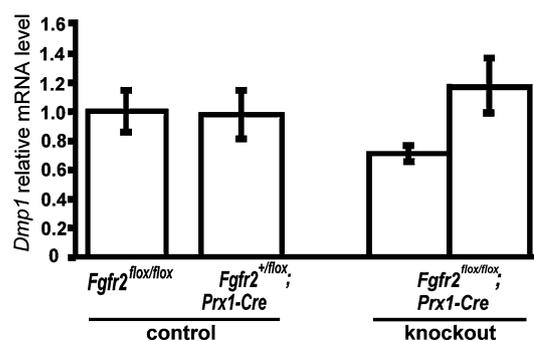


Fig. 10 *Fgfr2* inactivation does not affect *Dmp1* mRNA expression in vivo. *Dmp1* expression in the combined samples of tibiae and femora of E18.5 *Fgfr2*^{flox/flox}; *Prx1-Cre* and control littermate embryos was examined by real-time PCR. Data represent mean \pm standard deviations of samples from each individual embryo

treated with 2 μ g/ml FGF23 and 2 μ g/ml Klotho, the co-receptor for FGF23, either alone or in combination in the presence of 50 μ g/ml heparin. The combined treatment of FGF23 and Klotho upregulated *Dmp1* mRNA expression fourfold at 8 h after the treatment, while either FGF23 or Klotho alone did not affect *Dmp1* expression (Fig. 9).

ERK1 and ERK2 inactivation abolishes *Dmp1* expression in vivo

To examine the role of *Fgfr2* and ERK1/ERK2 in *Dmp1* expression in vivo, we examined *Dmp1* expression in the skeletal elements, in which *Fgfr2* and ERK1/ERK2 are inactivated using the *Prx1-Cre* transgene. While *Dmp1* was normally expressed in the skeletal elements of *Fgfr2*^{flox/flox}; *Prx1-Cre* embryos (Fig. 10), *Dmp1* expression was remarkably decreased in the humeri and tibiae of *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* embryos at embryonic day 16.5 (E16.5) (Fig. 11). In the humeri and tibiae, *Dmp1* expression of *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice was about 1.0 and 10% of control littermates, respectively.

We further examined *Dmp1* protein expression in osteocytes of *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice by immunohistochemistry. Although *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice showed a remarkable impairment of bone formation, bone-like architecture was observed in the diaphyses of long bones, and osteocyte-like cells were found within the bone-like matrix (Fig. 12). While intense staining for *Dmp1* was observed in osteocytes and their surrounding matrices in control mice, immunoreactivity for *Dmp1* was remarkably reduced in *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice. Collectively, these observations indicate that ERK1 and ERK2 are essential for *Dmp1* expression in osteocytes in vivo.

Since *Dmp1* is involved in phosphate regulation through FGF23, we further examined serum phosphate concentrations of *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice at P0.

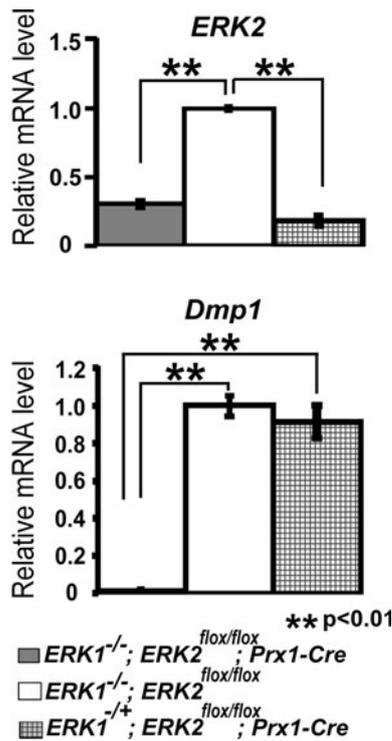


Fig. 11 *ERK1* and *ERK2* inactivation abolishes *Dmp1* mRNA expression in vivo. *ERK2* and *Dmp1* expression in the humeri of E16.5 *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* and control littermate embryos was examined by real-time PCR. Data represent mean ± standard deviations

There were no statistically significant differences between *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice and control mice (data not shown).

ERK1 and *ERK2* inactivation disrupts formation of lacunar-canalicular system

To examine the effects of *ERK1* and *ERK2* inactivation on the development of lacunar–canalicular system, acid-etched, resin-casted bone samples of *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice were further examined by scanning electron microscopy. While osteocytes in the control mice show a typical morphology with dendritic processes, osteocytes in *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice lack dendritic processes, indicating that *ERK1* and *ERK2* inactivation disrupts the formation of the osteocyte lacunar-canalicular system (Fig. 13).

Normal bone mineralization in the absence of *ERK1* and *ERK2*

Since FGF signaling regulates mineralization-related genes in osteoblasts and osteocytes, we assessed matrix mineralization in the skeletal elements of *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice by von Kossa staining at E16.5 and Goldner’s trichrome staining at P0 (Fig. 14 and data not shown). We observed no differences in matrix mineralization

Fig. 12 Immunohistochemical analysis for *Dmp1* protein expression in the tibiae of *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice at postnatal day 0. The staining for *Dmp1* was remarkably reduced in osteocytes (blue arrows) of *ERK1*^{-/-}; *ERK2*^{flox/flox}; *Prx1-Cre* mice, whereas control littermate *ERK1*^{-/-}; *ERK2*^{flox/flox} mice showed intense staining in osteocytes and their surrounding matrices. The boxed area in the upper panel is magnified in the bottom panel

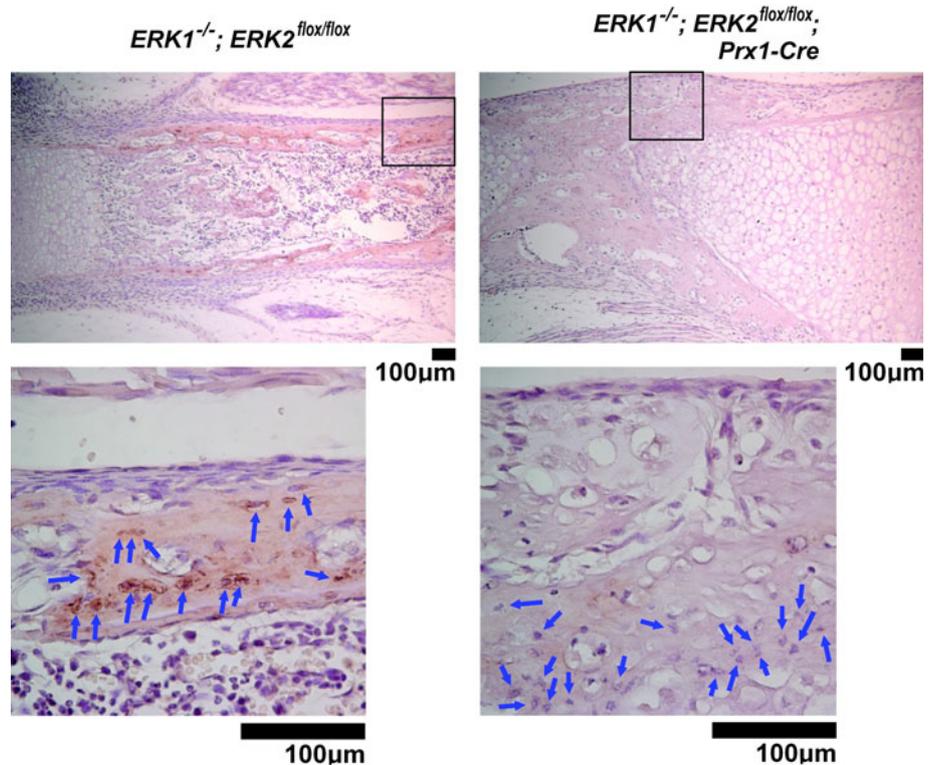


Fig. 13 Scanning electron microscopy images of the acid-etched, resin-casted osteocyte lacunar–canalicular system in the tibia at postnatal day 0. While control $ERK1^{+/-}$; $ERK2^{flox/flox}$; $Prx1-Cre$ mice show osteocytes with characteristic dendritic processes and organized canaliculi (left panel), osteocytes in $ERK1^{-/-}$; $ERK2^{flox/flox}$; $Prx1-Cre$ mice lack dendritic processes, and no canalicular system is observed

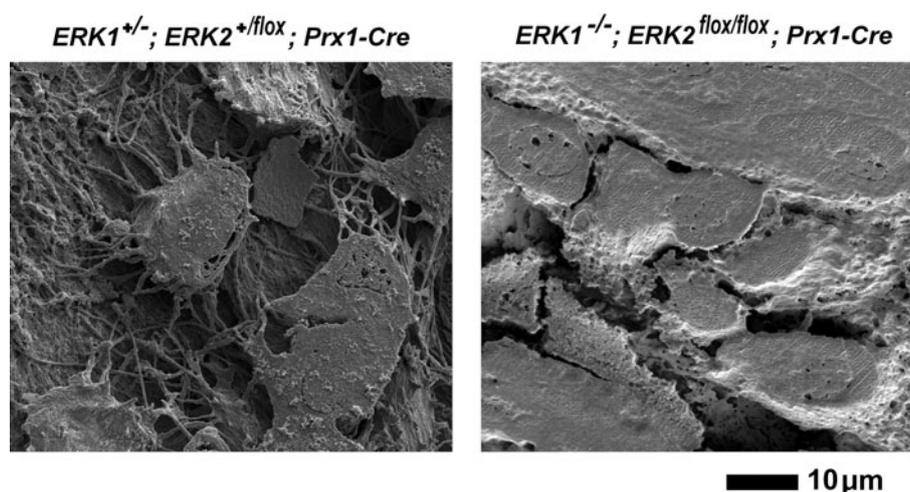
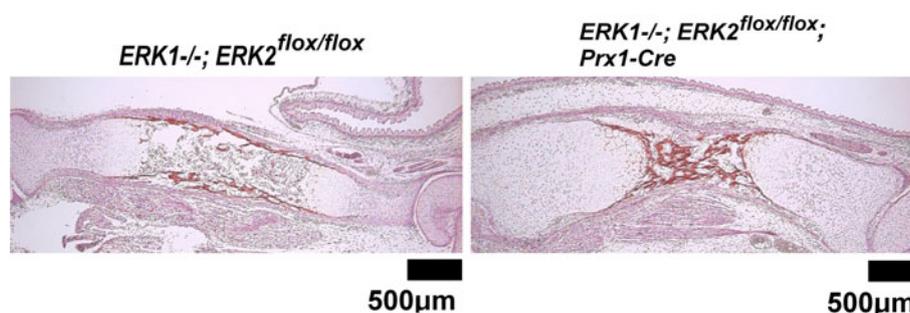


Fig. 14 von Kossa Staining of the tibia of $ERK1^{-/-}$; $ERK2^{flox/flox}$; $Prx1-Cre$ and control embryos at E16.5. No difference in matrix mineralization was observed between $ERK1^{-/-}$; $ERK2^{flox/flox}$; $Prx1-Cre$ and control mice



between control mice and $ERK1^{-/-}$; $ERK2^{flox/flox}$; $Prx1-Cre$ mice. This observation suggests that despite impaired osteoblast and osteocyte differentiation in $ERK1^{-/-}$; $ERK2^{flox/flox}$; $Prx1-Cre$ mice, the mineralization of bone matrix proceeds in the absence of ERK1 and ERK2.

Discussion

Our microarray analysis in MLO-Y4 osteocytes indicated that a number of mineralization-related genes are regulated by FGF2 via the ERK MAPK pathway. *Dmp1* is a critical regulator of skeletal mineralization and phosphate homeostasis. *Slc20a1* (Pit-1) is a sodium-dependent Pi cotransporter that regulates phosphate homeostasis [27]. Matrix Gla protein (MGP) is an inhibitor of mineralization implicated in vessel and cartilage calcification [28]. *Ank* is a transmembrane protein that transports intracellular pyrophosphate to the extracellular milieu [29]. *Enpp1* (ectonucleotide pyrophosphatase/phosphodiesterase, PC-1) is a major generator of extracellular pyrophosphate (PPi), which inhibits hydroxyapatite crystal deposition and growth [30]. All of these genes were upregulated by FGF2, and U0126 inhibited FGF2-induced upregulation. While

FGF2 regulation of *Enpp1* and *Ank* has been reported in primary calvarial pre-osteoblasts [31, 32], the upregulation of *Mgp*, *Slc20a1*, and *Dmp1* by FGFs has not been described previously. These results indicate that FGF signaling coordinately regulates mineralization-related genes in osteoblasts and osteocytes. Furthermore, the basal expression of *Ank*, *Slc20a1*, *Enpp1*, and *Dmp1* was inhibited by the selective FGF receptor inhibitor PD173074 in mouse calvaria maintained in organ culture, strongly suggesting that FGF signaling is essential for the normal expression of *Ank*, *Slc20a1*, *Enpp1*, and *Dmp1*. While the significance of the coordinated regulation requires further investigation, the increase in *Ank*, *Mgp*, *Enpp1* expression by FGF2 is consistent with previous reports showing the inhibitory effects of FGF signaling on mineralization in differentiated osteoblasts [3, 31, 33, 34]. While our experiments in primary calvaria osteoblasts also showed inhibitory effects of FGF signaling on mineralization, positive effects of FGF and ERK signaling on mineralization have been reported in other systems [35]. The apparent discrepancy may be due to the roles of FGF and ERK in promoting osteoblast differentiation, which in turn results in increased mineralization, while FGF and ERK also upregulate genes such as *Ank*, *Mgp*, and *Enpp1* that

inhibit mineralization. Depending on the specific experimental condition, either the former or the latter mechanisms may become more predominant.

Recent studies using genetically engineered mouse models have indicated that *Dmp1* plays an important role in skeletal mineralization, osteoblast-osteocyte transition, and osteocyte maturation [11, 15]. Our experiments indicate that FGF2 upregulates *Dmp1* expression in cells in the osteoblast lineage through the ERK MAPK pathway, and FGF and ERK signaling is required for *Dmp1* expression in osteoblasts and osteocytes. Since *Dmp1* is predominantly expressed in osteocytes in the osteoblast lineage, our results suggest that FGF and ERK signaling play an essential role in the late stages of osteoblast differentiation. The role of FGF on osteocyte differentiation may not be a simple acceleration, since we also found that FGF2 inhibits *Sost* expression in primary calvaria osteoblasts (data not shown).

Another important function of DMP1 is the regulation of phosphate homeostasis through FGF23. FGF23 is secreted from osteocytes, and circulating FGF23 increases renal phosphate excretion. Loss-of-function mutations in DMP1 cause ARHR in humans, and both patients with DMP1 mutations and *Dmp1*-null mice show hypophosphatemia with increased FGF23 expression in osteocytes. We found that FGF23 increases *Dmp1* expression in MLO-Y4 cells in the presence of Klotho, a cofactor essential for FGF23 receptor binding [36, 37]. The lack of response to FGF23 alone is consistent with our observation that *Klotho* mRNA was not detected in MLO-Y4 cells by real-time PCR (data not shown). Our results suggest the possibility that osteocyte-derived FGF23 might regulate *Dmp1* expression locally depending on the availability of Klotho. We also examined *Fgf23* expression in FGF2- and U0126-treated MLO-Y4 cells. We found that FGF23 expression was not affected by FGF2 or U0126 in MLO-Y4 cells, indicating that FGF2 and U0126 regulate *Dmp1* expression independent of *Fgf23* expression (data not shown). Our results also showed that the regulation of *Dmp1* expression by FGF2 is modulated by extracellular inorganic phosphate levels. It is interesting to speculate that FGF and inorganic phosphate coordinately regulate *Dmp1* expression in osteocytes to regulate phosphate homeostasis.

FGF receptors are likely to play both redundant and distinct roles in bone development, since various mutations in FGF receptors in mice and humans cause both overlapping and distinct phenotypes [2, 3, 38, 39]. Given the importance of FGF signaling and *Dmp1* in the skeletal system, it is of interest to identify FGF receptors that regulate *Dmp1* expression. PD173074, which blocks all isoforms of FGF receptors, inhibited *Dmp1* expression in the mouse calvaria, indicating the requirement of FGF signaling for *Dmp1* expression. However, *Dmp1* was

normally expressed in the skeletal elements of *Fgfr2^{fllox/fllox}; Prx1-Cre* mice, suggesting that signals from other FGF receptors contribute to *Dmp1* expression (Fig. 10). This is also consistent with our observations that FGF2 upregulates *Dmp1* in MLO-Y4 cells that express only *Fgfr1* and *Fgfr3*. The remarkable inhibition of *Dmp1* expression by U0126 in MLO-Y4 cells and the lack of *Dmp1* expression in osteocytes of *ERK1^{-/-}; ERK2^{fllox/fllox}; Prx1-Cre* mice indicate the requirement of ERK MAPK signaling for *Dmp1* expression. Because ERK1 and ERK2 are common downstream effectors of Fgfr1, Fgfr2, Fgfr3, and Fgfr4, signals originating from any of the four FGF receptors may regulate *Dmp1*.

In summary, we have shown that a number of mineralization-related genes are regulated by FGF2 in a MAPK-dependent manner in the osteocyte cell line MLO-Y4. Our experiments also indicate that *Dmp1* expression in osteoblasts and osteocytes requires FGF and ERK MAPK signaling, and ERK1 and ERK2 are essential for osteocyte differentiation and the development of the osteocyte lacunar-canalicular system. Further analysis of FGF and MAPK signaling in osteocytes will provide novel information on the mechanisms of osteocyte differentiation and mineral homeostasis.

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References

1. Montero A, Okada Y, Tomita M, Ito M, Tsurukami H, Nakamura T, Doetschman T, Coffin JD, Hurley MM (2000) Disruption of the fibroblast growth factor-2 gene results in decreased bone mass and bone formation. *J Clin Invest* 105:1085–1093
2. Yu K, Xu J, Liu Z, Susic D, Shao J, Olson EN, Towler DA, Ornitz DM (2003) Conditional inactivation of FGF receptor 2 reveals an essential role for FGF signaling in the regulation of osteoblast function and bone growth. *Development* 130:3063–3074
3. Jacob AL, Smith C, Partanen J, Ornitz DM (2006) Fibroblast growth factor receptor 1 signaling in the osteo-chondrogenic cell lineage regulates sequential steps of osteoblast maturation. *Dev Biol* 296:315–328
4. Wilkie AO (2005) Bad bones, absent smell, selfish testes: the pleiotropic consequences of human FGF receptor mutations. *Cytokine Growth Factor Rev* 16:187–203
5. Debais F, Lemonnier J, Hay E, Delannoy P, Caverzasio J, Marie PJ (2001) Fibroblast growth factor-2 (FGF-2) increases N-cadherin expression through protein kinase C and Src-kinase pathways in human calvaria osteoblasts. *J Cell Biochem* 81:68–81
6. Murakami S, Balmes G, McKinney S, Zhang Z, Givol D, de Crombrughe B (2004) Constitutive activation of MEK1 in

- chondrocytes causes Stat1-independent achondroplasia-like dwarfism and rescues the Fgfr3-deficient mouse phenotype. *Genes Dev* 18:290–305
7. Matsushita T, Wilcox WR, Chan YY, Kawanami A, Bükülmez H, Balmes G, Krejci P, Mekikian PB, Otani K, Yamaura I, Warman ML, Givol D, Murakami S (2009) FGFR3 promotes synchondrosis closure and fusion of ossification centers through the MAPK pathway. *Hum Mol Genet* 18:227–240
 8. Matsushita T, Chan YY, Kawanami A, Balmes G, Landreth GE, Murakami S (2009) Extracellular signal-regulated kinase 1 (ERK1) and ERK2 play essential roles in osteoblast differentiation and in supporting osteoclastogenesis. *Mol Cell Biol* 29:5843–5857
 9. Bentires-Alj M, Kontaridis MI, Neel BG (2006) Stops along the RAS pathway in human genetic disease. *Nat Med* 12:283–285
 10. Aoki Y, Niihori T, Narumi Y, Kure S, Matsubara Y (2008) The RAS/MAPK syndromes: novel roles of the RAS pathway in human genetic disorders. *Hum Mutat* 29:992–1006
 11. Feng JQ, Ward LM, Liu S, Lu Y, Xie Y, Yuan B, Yu X, Rauch F, Davis SI, Zhang S, Rios H, Drezner MK, Quarles LD, Bonewald LF, White KE (2006) Loss of DMP1 causes rickets and osteomalacia and identifies a role for osteocytes in mineral metabolism. *Nat Genet* 38:1310–1315
 12. Tatsumi S, Ishii K, Amizuka N, Li M, Kobayashi T, Kohno K, Ito M, Takeshita S, Ikeda K (2007) Targeted ablation of osteocytes induces osteoporosis with defective mechanotransduction. *Cell Metab* 5:464–475
 13. Xiao ZS, Quarles LD (2010) Role of the polycystin-primary cilia complex in bone development and mechanosensing. *Ann N Y Acad Sci* 1192:410–421
 14. Dallas SL, Bonewald LF (2010) Dynamics of the transition from osteoblast to osteocyte. *Ann N Y Acad Sci* 1192:437–443
 15. Lu Y, Yuan B, Qin C, Cao Z, Xie Y, Dallas SL, McKee MD, Drezner MK, Bonewald LF, Feng JQ (2011) The biological function of DMP1 in osteocyte maturation is mediated by its 57 kDa C-terminal fragment. *J Bone Miner Res* 26:331–340
 16. Kato Y, Windle JJ, Koop BA, Mundy GR, Bonewald LF (1997) Establishment of an osteocyte-like cell line, MLO-Y4. *J Bone Miner Res* 12:2014–2023
 17. Hill TP, Später D, Taketo MM, Birchmeier W, Hartmann C (2005) Canonical Wnt/beta-catenin signaling prevents osteoblasts from differentiating into chondrocytes. *Dev Cell* 8:727–738
 18. Selcher JC, Nekrasova T, Paylor R, Landreth GE, Sweatt JD (2001) Mice lacking the ERK1 isoform of MAP kinase are unimpaired in emotional learning. *Learn Mem* 8:11–19
 19. Samuels IS, Karlo JC, Faruzzi AN, Pickering K, Herrup K, Sweatt JD, Saitta SC, Landreth GE (2008) Deletion of ERK2 mitogen-activated protein kinase identifies its key roles in cortical neurogenesis and cognitive function. *J Neurosci* 28:6983–6995
 20. Logan M, Martin JF, Nagy A, Lobe C, Olson EN, Tabin CJ (2002) Expression of Cre Recombinase in the developing mouse limb bud driven by a Prxl enhancer. *Genesis* 33:77–80
 21. Kubek DJ, Gattone VH 2nd, Allen MR (2010) Methodological assessment of acid-etching for visualizing the osteocyte lacunar-canalicular networks using scanning electron microscopy. *Micro Res Tech* 73:182–186
 22. Koziczak M, Holbro T, Hynes NE (2004) Blocking of FGFR signaling inhibits breast cancer cell proliferation through down-regulation of D-type cyclins. *Oncogene* 23:3501–3508
 23. Pardo OE, Latigo J, Jeffery RE, Nye E, Poulosom R, Spencer-Dene B, Lemoine NR, Stamp GW, Aboagye EO, Seckl MJ (2009) The fibroblast growth factor receptor inhibitor PD173074 blocks small cell lung cancer growth in vitro and in vivo. *Cancer Res* 69:8645–8651
 24. St-Germain JR, Taylor P, Tong J, Jin LL, Nikolic A, Stewart II, Ewing RM, Dharsee M, Li Z, Trudel S, Moran MF (2009) Multiple myeloma phosphotyrosine proteomic profile associated with FGFR3 expression, ligand activation, and drug inhibition. *Proc Natl Acad Sci USA* 106:20127–20132
 25. Kato Y, Boskey A, Spevak L, Dallas M, Hori M, Bonewald LF (2001) Establishment of an osteoid preosteocyte-like cell MLO-A5 that spontaneously mineralizes in culture. *J Bone Miner Res* 16:1622–1633
 26. Qin C, Brunn JC, Cook RG, Orkiszewski RS, Malone JP, Veis A, Butler WT (2003) Evidence for the proteolytic processing of dentin matrix protein 1. Identification and characterization of processed fragments and cleavage sites. *J Biol Chem* 278:34700–34708
 27. Li X, Giachelli CM (2007) Sodium-dependent phosphate cotransporters and vascular calcification. *Curr Opin Nephrol Hypertens* 16:325–328
 28. Luo G, Ducy P, McKee MD, Pinero GJ, Loyer E, Behringer RR, Karsenty G (1997) Spontaneous calcification of arteries and cartilage in mice lacking matrix GLA protein. *Nature* 386:78–81
 29. Ho AM, Johnson MD, Kingsley DM (2000) Role of the mouse ank gene in control of tissue calcification and arthritis. *Science* 289:265–270
 30. Hesse L, Johnson KA, Anderson HC, Narisawa S, Sali A, Goding JW, Terkeltaub R, Millan JL (2002) Tissue-nonspecific alkaline phosphatase and plasma cell membrane glycoprotein-1 are central antagonistic regulators of bone mineralization. *Proc Natl Acad Sci USA*. 99:9445–9449
 31. Hatch NE, Nociti F, Swanson E, Bothwell M, Somerman M (2005) FGF2 alters expression of the pyrophosphate/phosphate regulating proteins, PC-1, ANK and TNAP, in the calvarial osteoblastic cell line, MC3T3E1(C4). *Connect Tissue Res* 46:184–192
 32. Hatch NE, Franceschi RT (2009) Osteoblast differentiation stage-specific expression of the pyrophosphate-generating enzyme PC-1. *Cells Tissues Organs* 189:65–69
 33. Mansukhani A, Bellosta P, Sahni M, Basilico C (2000) Signaling by fibroblast growth factors (FGF) and fibroblast growth factor receptor 2 (FGFR2)-activating mutations blocks mineralization and induces apoptosis in osteoblasts. *J Cell Biol* 149:1297–1308
 34. Kono SJ, Oshima Y, Hoshi K, Bonewald LF, Oda H, Nakamura K, Kawaguchi H, Tanaka S (2007) Erk pathways negatively regulate matrix mineralization. *Bone* 40:68–74
 35. Miraoui H, Oudina K, Petite H, Tanimoto Y, Moriyama K, Marie PJ (2009) Fibroblast growth factor receptor 2 promotes osteogenic differentiation in mesenchymal cells via ERK 1/2 and protein kinase C signaling. *J Biol Chem* 284:4897–4904
 36. Kurosu H, Ogawa Y, Miyoshi M, Yamamoto M, Nandi A, Rosenblatt KP, Baum MG, Schiavi S, Hu MC, Moe OW, Kuro-o M (2006) Regulation of fibroblast growth factor-23 signaling by klotho. *J Biol Chem* 281:6120–6123
 37. Urakawa I, Yamazaki Y, Shimada T, Iijima K, Hasegawa H, Okawa K, Fujita T, Fukumoto S, Yamashita T (2006) Klotho converts canonical FGF receptor into a specific receptor for FGF23. *Nature* 444:770–774
 38. Bellus GA, Gaudenz K, Zackai EH, Clarke LA, Szabo J, Francomano CA, Muenke M (1996) Identical mutations in three different fibroblast growth factor receptor genes in autosomal dominant craniosynostosis syndromes. *Nat Genet* 14:174–176
 39. White KE, Cabral JM, Davis SI, Fishburn T, Evans WE, Ichikawa S, Fields J, Yu X, Shaw NJ, McLellan NJ, McKeown C, Fitzpatrick D, Yu K, Ornitz DM, Econs MJ (2005) Mutations that cause osteoglyphonic dysplasia define novel roles for FGFR1 in bone elongation. *Am J Hum Genet* 76:361–367